

A novel mutation assay with a non-selective protocol using a next-generation DNA sequencer

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GENOTOXICITY TESTS

In vitro tests

(using cell culture or bacteria)

1. Rec assay
 2. *umu* test
 3. **Reverse mutation assay**
(Ames test)
 5. Forward mutation assay
 6. Unscheduled DNA synthesis test
 5. Sister chromatic exchange test
 6. Chromosomal aberration test
 7. Micronucleus assay
 8. Comet assay
- etc.

In vivo tests

(using animals)

1. Micronucleus assay
 2. Sister chromatic exchange test
 3. Chromosomal aberration test
 4. Unscheduled DNA synthesis test
 5. *Pig-a* assay
 6. Transgenic animal gene mutation assay
 7. Drosophila Spot test
 8. Comet assay
- etc.

GENOTOXICITY TEST Conventional method

Ames test

- Data can be available within a week
- No need of special facilities
- Reproducible and reliable data
- Accumulation of reliable data in accordance with GLP
- ◆ Data are the number of colonies

GENOTOXICITY TEST

From a different perspective



Identification of whether the chemical is hazardous is the principal aim of Ames test.



We can directly detect the mutation induced by the chemical in the genome with **NGS**



A **N**EXT **G**ENERATION **S**EQUENCER MUST BE USEFUL !

GENOTOXICITY TEST

Novel method

Whole genome sequencing

- Data can be available within a week ✓
- The special facility is required ↓
- Reproducible and stable data ✓
- No data at present ↓
- ◆ Data is **ACTUAL** base changes !

COMPARISON

| <u>Ames test</u> | <u>Whole genome sequencing</u> |
|--|--|
| 5 strains and 4 or more doses | One strain is enough and not many doses are needed |
| Frameshift or base substitution | Any type of mutation |
| Minimal plates for two-night incubation | LB plates for one night incubation |
| His ⁺ selection | Non-selective method |
| 5000 µg/plate for a top dose, and one hundred plates | Lower doses, ~10 plates |
| Precipitation or killing effect of samples may disturb the assay | Colony formation is minimum requirement |

PROCEDURE

Conventional Ames test

Test chemical: AF-2 (0.01 μ g/plate)
DMSO as a solvent control
Strain: TA100 , without S9mix



Randomly pick up the revertants
Few clones for each condition
Prepare the genomic DNA

Whole genome sequencing of
the clones using an NGS, Miseq

The expected base substitutions at the target site of His⁺ reversion

| Treatment | Clone ID | Frequency (%) | Coverage | Gene | Site | Mutation | Amino Acid Change |
|-----------|----------|---------------|----------|-------------|------|----------|-------------------|
| DMSO | 1 | 100 | 14 | <i>hisG</i> | 206 | C>A | Pro69His |
| | 2 | 100 | 17 | <i>hisG</i> | 205 | C>A | Pro69Ile |
| | 3 | 100 | 14 | <i>hisG</i> | 206 | C>A | Pro69His |
| | 4 | 100 | 14 | <i>hisG</i> | 205 | C>A | Pro69Ile |
| | 5 | 100 | 17 | <i>hisG</i> | 205 | C>T | Pro69Phe |
| AF-2 | 6 | 100 | 29 | <i>hisG</i> | 206 | C>A | Pro69His |
| | 7 | 100 | 15 | <i>hisG</i> | 206 | C>A | Pro69His |
| | 8 | 100 | 37 | <i>hisG</i> | 206 | C>T | Pro69Leu |
| | 9 | 100 | 10 | <i>hisG</i> | 205 | C>T | Pro69Phe |
| | 10 | 100 | 26 | <i>hisG</i> | 205 | C>G | Pro69Val |

RESULTS - 1

Base Substitutions Induced on the Whole Genome of the Ames tester Strain TA100 Treated with AF-2

| Treatment | Clone ID | Frequency (%) | Coverage | Gene | Site | Mutation | Seq. context | Amino acid change |
|-----------|----------|---------------|----------|-------------|------|----------|--------------|-------------------|
| DMSO | 2 | 100 | 31 | <i>hypE</i> | 831 | T>A | GAT[T]GCC | |
| AF-2 | 6 | 100 | 29 | <i>yraP</i> | 25 | G>A | GCA[G]TCC | V 19 I |
| | 8 | 96.7 | 30 | <i>cueO</i> | 964 | C>T | CCG[C]TGC | |
| | 8 | 100 | 29 | <i>ybiR</i> | 832 | C>A | GCA[C]TGT | L278M |
| | 8 | 100 | 38 | <i>xseA</i> | 996 | C>G | GGC[C]AGG | |
| | 10 | 100 | 27 | <i>hisG</i> | 231 | G>C | GCT[G]GAA | |

RESULTS - 2

ENU treated strain YG7108*

Induced Reverse Mutations in Ames Test and Base Substitutions Induced in the Whole Genome

| ENU µg/plate | Ames test revertants/plate | Whole genome sequence (per genome) | | | |
|-----------------|-------------------------------|---------------------------------------|------------|------------|------------|
| | | total | G:C to A:T | A:T to G:C | others |
| 0 | 17 | 0 | 0 | 0 | 0 |
| | | 0 | 0 | 0 | 0 |
| | | 0 | 0 | 0 | 0 |
| | | 0 | 0 | 0 | 0 |
| 50 | 3,250 | 12 | 12 | 0 | 0 |
| | | 19 | 16 | 2 | 1(GCtoCG) |
| | | 14 | 14 | 0 | 0 |
| | | 8 | 8 | 0 | 0 |
| 250 | 8,953 | 64 | 63 | 1 | 0 |
| | | 62 | 57 | 4 | 1(GCtoTA) |
| | | 48 | 44 | 4 | |
| | | 71 | 65 | 5 | 1(Gcto CG) |

ADVANCED PROTOCOL

No selection!

- I. Treatment with the test chemical is conducted in liquid medium.
- II. The treated culture is spread onto an LB plate after appropriate dilution.
- III. Colonies are obtained without phenotypic selection.
- IV. Genomic analysis with a next-generation DNA sequencer is performed for a few randomly chosen colonies.

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RESULTS - 3

Mutation spectrum of the genome in TA1535 treated with ENU with no selection

| Pre-incubation time (min) | x 10 ⁶ cells | Mutation/genome | Ave. | GC to AT | GC to TA | GC to CG | AT to GC | AT to CG | AT to TA | Insertion | Deletion |
|---------------------------|-------------------------|-----------------|------|----------|----------|----------|----------|----------|----------|-----------|----------|
| 0 | 295 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | | 0 | | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | | 0 | | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 20 | 47 | 19 | 22 | 16 | 0 | 0 | 3 | 0 | 0 | 0 | 0 |
| | | 18 | | 17 | 0 | 0 | 1 | 0 | 0 | 0 | 0 |
| | | 29 | | 27 | 0 | 0 | 1 | 0 | 0 | 0 | 0 |

| | | | | | | | | | | | |
|----|----|----|----|----|---|---|---|---|---|---|---|
| 40 | 46 | 4 | 9 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | | 22 | | 20 | 0 | 0 | 1 | 0 | 0 | 0 | 1 |
| | | 1 | | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 60 | 43 | 16 | 16 | 16 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| | | 9 | | 6 | 0 | 0 | 2 | 0 | 1 | 0 | 0 |
| | | 23 | | 21 | 0 | 0 | 2 | 0 | 0 | 0 | 0 |

CONCLUSION

Whole genome sequencing would realize

THAT

**ANY MUTATIONS ! can be detected
in ONE STRAIN !**

with MORE PRECISE DATA !

at LESS COST !

IN THE FUTURE

- Considering the remarkable recent advances in the performance of DNA sequencers, it would soon be possible to easily determine whole genomes of rodents or cultured human cells exposed to chemicals.
- However, taking into consideration of the principal aim of Ames test, that is, determining of hazard identification on DNA induced by chemicals, bacteria still have an advantage due to its small size of genome and identical composition of DNA to the other organisms.