
1 NOTE ON THE MONOGRAPH

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3 *This monograph has been thoroughly revised further to the contamination events in*
4 *2008 to ensure appropriate quality control for unfractionated heparin. The style and*
5 *presentation have also been updated in line with the current version of the Style guide.*

6 **Definition:** *the minimum potency limit has been raised after an enquiry among*
7 *European manufacturers regarding the quality of currently marketed heparin batches;*
8 *only 1 grade of heparin has been kept as the present 2-tiered specification no longer*
9 *reflects the situation in Europe.*

10
11 **Production:** *the tests for nuclear magnetic resonance spectrometry (NMR) and capillary*
12 *electrophoresis previously introduced in the 1st-step revision applicable from 1 August*
13 *2008 have been deleted, as detailed tests are now provided under Identification*
14 *and Tests; statements have been added to emphasise the need for a reliable quality*
15 *management system throughout production and, based on current practice among*
16 *European manufacturers, for confirming the identity of the source species as well as*
17 *the absence of any material issued from other species likely to contaminate the drug*
18 *substance. This monograph is also revised to harmonise the information related to*
19 *the source species for substances of human and animal origin and its presentation*
20 *in monographs. The statement relative to the origin of the substance is moved under*
21 *Definition accordingly and a paragraph is added regarding the health of the animals*
22 *used for the preparation of heparin calcium.*

23 **Identification:** *the tests for specific optical rotation and zone electrophoresis have*
24 *been replaced by the highly specific ¹H-NMR and strong anion-exchange liquid*
25 *chromatography (SAX-HPLC) tests; ¹H-NMR has been selected for its ability not only to*
26 *allow identification of heparin, but also to alert users to possible contaminations.*

27
28 **Nucleotidic impurities:** *the limit has been tightened, based on current batch data.*

29 **Protein:** *the Lowry test method has been introduced to replace the absorbance test.*

30
31 **Related substances:** *a SAX-HPLC-based test has been introduced, allowing the*
32 *differentiation of natural contaminants linked to the production process (such as*
33 *dermatan sulfate and chondroitin sulfate) from chemically synthesised contaminants;*
34 *a limit for the sum of dermatan sulfate and chondroitin sulfate, which co-elute in this*
35 *method, is proposed, further to consideration of current batch data.*

36 **Nitrogen:** *a lower limit has been added, based on current batch data.*

37
38 **Heavy metals:** *method C has been replaced by method F, in line with the general policy*
39 *for heavy metals tests.*

40 **Sulfated ash:** *in view of the highly specific tests introduced into the monograph, this*
41 *test has become redundant and has therefore been deleted.*

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43 08/2010:0332

44
45 **HEPARIN CALCIUM**

46
47 Heparinum calcicum

1 DEFINITION

2 Preparation containing the calcium salt of a sulfated glycosaminoglycan present in
3 mammalian tissues. It is prepared either from the lungs of cattle or from the intestinal
4 mucosae of pigs, cattle or sheep. On complete hydrolysis, it liberates D-glucosamine,
5 D-glucuronic acid, L-iduronic acid, acetic acid and sulfuric acid. It has the property of
6 delaying the clotting of blood.
7

8 *Potency*: minimum 180 IU/mg (dried substance).
9

10 PRODUCTION

11 The animals from which heparin calcium is derived must fulfil the requirements for the
12 health of animals suitable for human consumption. All stages of production and sourcing
13 are subjected to a suitable quality management system. The identity of the source species
14 and the absence of material from the other species is verified by appropriate testing
15 during production.
16

17 It is produced by methods of manufacturing designed to minimise or eliminate substances
18 lowering blood pressure.

19 CHARACTERS

20 *Appearance*: white or almost white, hygroscopic powder.

21 *Solubility*: freely soluble in water.
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24 IDENTIFICATION

25 A. It delays the clotting of recalcified citrated sheep plasma (see Assay).
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28 B. Nuclear magnetic resonance spectrometry (2.2.33).
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30 *Preparation*: dissolve 20 mg of the substance to be examined in 0.7 mL of a 20 µg/mL
31 solution of *deuterated sodium trimethylsilylpropionate R* in *deuterium oxide R*.

32 *Comparison*: dissolve 20 mg of *heparin calcium for NMR identification CRS* in
33 0.7 mL of a 20 µg/mL solution of *deuterated sodium trimethylsilylpropionate R* in
34 *deuterium oxide R*.

35 *Apparatus*: spectrometer operating at minimum 300 MHz.
36

37 *Acquisition of ¹H-NMR spectra*:

- 38 – *number of transients*: minimum 16; it is adjusted until the signal-to-noise ratio is at
39 least 1000:1 for the heparin methyl signal at 2.04 ppm;
- 40 – *temperature*: about 25 °C; test sample and reference spectra have to be obtained at
41 the same temperature;
- 42 – *acquisition time*: minimum 2 s;
- 43 – *repetition time* (acquisition time plus delay): minimum 4 s;
- 44 – *spectral width*: 10-12 ppm, centred at around 4.5 ppm;
- 45 – *pulse width*: to give a flip angle between 30° and 90°.
46
47

1 *Processing:*

- 2 – *exponential line-broadening window function*: 0.3 Hz;
3
4 – Fourier transformation;
5 – trimethylsilylpropionate reference signal set at 0.00 ppm.

6 *Results:*

- 7 – the large heparin calcium signals must be present: 2.05 ppm, 3.29 ppm (doublet),
8 4.37 ppm, 5.35 ppm and 5.43 ppm, all within ± 0.03 ppm;
9 – the $^1\text{H-NMR}$ spectrum obtained with the test sample and that obtained with *heparin*
10 *calcium for NMR identification CRS* are compared qualitatively after the 2 spectra
11 have been normalised so as to have a similar intensity; dermatan sulfate with a
12 methyl signal at 2.08 ± 0.02 ppm may be observed; no unidentified signals larger
13 than 4 per cent compared to the height of the heparin signal at 5.43 ppm are present
14 in the ranges 0.10-2.00 ppm, 2.10-3.10 ppm and 5.70-8.00 ppm; signals from the
15 solvent or process-related substances may be present and have to be identified to
16 be accepted.

- 17
18 C. Liquid chromatography (2.2.29) as described in the test for related substances with the
19 following modifications.

20 *Injection*: test solution (a) and reference solution (c).

21 *Relative retention* with reference to heparin (retention time = about 26 min): dermatan
22 sulfate and chondroitin sulfate = about 0.9; over-sulfated chondroitin sulfate =
23 about 1.3.

24 *System suitability*: reference solution (c):

- 25 – *peak-to-valley ratio*: minimum 1.3, where H_p = height above the baseline of the peak
26 due to dermatan sulfate + chondroitin sulfate and H_v = height above the baseline of
27 the lowest point of the curve separating this peak from the peak due to heparin.

28 *Results*: the principal peak in the chromatogram obtained with test solution (a) is
29 similar in retention time and shape to the principal peak in the chromatogram obtained
30 with reference solution (c).
31

- 32 D. It gives the reactions of calcium (2.3.1).

33 TESTS

34 **Appearance of solution.** The solution is clear (2.2.1) and not more intensely coloured
35 than intensity 5 of the range of reference solutions of the most appropriate colour (2.2.2,
36 *Method II*).

37
38 Dissolve a quantity equivalent to 50 000 IU in *water R* and dilute to 10 mL with the
39 same solvent.

40 **pH** (2.2.3): 5.5 to 8.0.

41 Dissolve 0.1 g in *carbon dioxide-free water R* and dilute to 10 mL with the same solvent.

42 **Nucleotidic impurities.** Dissolve 40 mg in 10 mL of *water R*. The absorbance (2.2.25)
43 measured at 260 nm is not greater than 0.15.

44 **Protein**: maximum 0.5 per cent (dried substance).

45 *Solution A.* Mix 2 volumes of a 10 g/L solution of *sodium hydroxide R* and 2 volumes of a
46 50 g/L solution of *sodium carbonate R* and dilute to 5 volumes with *water R*.
47

1 *Solution B.* Mix 2 volumes of a 12.5 g/L solution of *copper sulfate R* and 2 volumes of a
2 29.8 g/L solution of *sodium tartrate R* and dilute to 5 volumes with *water R*.

3 *Solution C.* Mix 1 volume of solution B and 50 volumes of solution A.

4
5 *Solution D.* Dilute a phosphomolybdotungstic reagent⁽¹⁾ 2- to 4-fold in *water R*. Suitable
6 dilutions produce solutions of pH 10.25 ± 0.25 after addition of solutions C and D to
7 the test and reference solutions.

8 *Test solution.* Dissolve the substance to be examined in *water R* to obtain a concentration
9 of 5 mg/mL.

10 *Reference solutions.* Dissolve *bovine albumin R* in *water R* to obtain a concentration of
11 100 mg/mL. Prepare dilutions of the solution in *water R* as prescribed in general chapter
12 2.5.33, method 2.

13 *Blank: water R.*

14
15 *Procedure.* To 1 mL of each reference solution, of the test solution and of the blank, add
16 5 mL of solution C. Allow to stand for 10 min. Add 0.5 mL of solution D, mix and allow
17 to stand at room temperature for 30 min. Determine the absorbances (2.2.25) of the
18 solutions at 750 nm, using the solution prepared from the blank as compensation liquid.

19 *Calculations.* As prescribed in general chapter 2.5.33, method 2.

20
21 **Related substances.** Liquid chromatography (2.2.29). *Reference solutions are stable at*
22 *room temperature for 24 h.*

23 *Test solution (a).* Dissolve an accurately weighed quantity of about 50 mg of the substance
24 to be examined in 5.0 mL of *water for chromatography R*. Mix using a vortex mixer until
25 dissolution is complete.

26
27 *Test solution (b).* Dissolve an accurately weighed quantity of about 0.1 g of the substance
28 to be examined in 1.0 mL of *water for chromatography R*. Mix using a vortex mixer until
29 dissolution is complete. Mix 500 μL of the solution and 250 μL of 1 M *hydrochloric acid*,
30 then add 50 μL of a 250 mg/mL solution of *sodium nitrite R*⁽²⁾. Mix gently and allow to
31 stand at room temperature for 40 min before adding 200 μL of 1 M *sodium hydroxide*
32 to stop the reaction.

33 *Reference solution (a).* Dissolve 250 mg of *heparin for physico-chemical analysis CRS*
34 in *water for chromatography R* and dilute to 2.0 mL with the same solvent. Mix using a
35 vortex mixer until dissolution is complete.

36
37 *Reference solution (b).* Add 1200 μL of reference solution (a) to 300 μL of *dermatan sulfate*
38 *and over-sulfated chondroitin sulfate CRS*. Mix using a vortex mixer to homogenise.

39 *Reference solution (c).* Add 100 μL of reference solution (b) to 900 μL of *water for*
40 *chromatography R*. Mix using a vortex mixer to homogenise.

41
42 *Reference solution (d).* Add 400 μL of reference solution (a) to 100 μL of *water for*
43 *chromatography R* and mix using a vortex mixer. Add 250 μL of 1 M *hydrochloric acid*,
44 then add 50 μL of a 250 mg/mL solution of *sodium nitrite R*. Mix gently and allow to
45 stand at room temperature for 40 min before adding 200 μL of 1 M *sodium hydroxide*
46 to stop the reaction.

47 (1) Folin-Ciocalteu's phenol reagent from Merck (reference 1.09001.0500) is suitable.

(2) Sodium nitrite, analytical reagent grade from Fischer scientific (batch 0886083) is suitable.

1 *Reference solution (e)*. To 500 μL of reference solution (b), add 250 μL of 1 M
 2 *hydrochloric acid*, then add 50 μL of a 250 mg/mL solution of *sodium nitrite R*. Mix
 3 gently and allow to stand at room temperature for 40 min before adding 200 μL of 1 M
 4 *sodium hydroxide* to stop the reaction.

5 *Precolumn:*

- 6 – size: $l = 0.05$ m, $\text{Ø} = 2$ mm;
- 7 – stationary phase: *anion exchange resin R* (13 μm)⁽³⁾.

9 *Column:*

- 10 – size: $l = 0.25$ m, $\text{Ø} = 2$ mm;
- 11 – stationary phase: *anion exchange resin R* (9 μm)⁽⁴⁾;
- 12 – temperature: 40 °C.

14 *Mobile phase:*

- 15 – *mobile phase A*: dissolve 0.40 g of *sodium dihydrogen phosphate R* in 1 L of *water for chromatography R* and adjust to pH 3.0 with *dilute phosphoric acid R*;
- 16 – *mobile phase B*: dissolve 0.40 g of *sodium dihydrogen phosphate R* in 1 L of *water for chromatography R*, add 140 g of *sodium perchlorate R*⁽⁵⁾ and adjust to pH 3.0 with *dilute phosphoric acid R*; filter and degas;

20	Time	Mobile phase A	Mobile phase B
21	(min)	(per cent V/V)	(per cent V/V)
22	0 - 10	75	25
23	10 - 35	75 \rightarrow 0	25 \rightarrow 100
24	35 - 40	0	100

25 *Flow rate*: 0.22 mL/min.

26 *Detection*: spectrophotometer at 202 nm.

27 *Equilibration*: at least 15 min.

28 *Injection*: 20 μL of test solution (b) and reference solutions (d) and (e).

29 *Relative retention* with reference to heparin (retention time = about 26 min): dermatan
 30 sulfate and chondroitin sulfate = about 0.9; over-sulfated chondroitin sulfate = about 1.3.

31 *System suitability:*

- 32 – the chromatogram obtained with reference solution (d) shows no peak at the retention
 33 time of heparin;
- 34 – *resolution*: minimum 3.0 between the peaks due to dermatan sulfate + chondroitin
 35 sulfate and over-sulfated chondroitin sulfate in the chromatogram obtained with
 36 reference solution (e).

37 *Limits:*

- 38 – *sum of dermatan sulfate and chondroitin sulfate*: not more than the area of the
 39 corresponding peak in the chromatogram obtained with reference solution (e) (2.0 per
 40 cent);
- 41 – *any other impurity*: no peaks other than the peak due to dermatan sulfate
 42 + chondroitin sulfate are detected.

43 (3) AG11-HC from Dionex (reference 052963) is suitable.

44 (4) AS11-HC from Dionex (reference 052961) is suitable.

45 (5) Normapur from VWR/Prolabo (reference 27988.232) is suitable.

1 **Nitrogen** (2.5.9): 1.5 per cent to 2.5 per cent (dried substance), determined on 0.100 g. |

2
3 **Calcium**: 9.5 per cent to 11.5 per cent (dried substance), determined on 0.200 g by
4 complexometric titration (2.5.11).

5 **Heavy metals** (2.4.8): maximum 30 ppm.

6
7 1.0 g complies with test F. Prepare the reference solution using 3.0 mL of *lead standard*
8 *solution* (10 ppm Pb) R. |

9 **Loss on drying** (2.2.32): maximum 8.0 per cent, determined on 1.000 g by drying at 60 °C
10 over *diphosphorus pentoxide* R at a pressure not exceeding 670 Pa for 3 h. —

11
12 **Bacterial endotoxins** (2.6.14): less than 0.01 IU per International Unit of heparin,
13 if intended for use in the manufacture of parenteral preparations without a further
14 appropriate procedure for the removal of bacterial endotoxins. The addition of divalent
15 cations may be necessary in order to fulfil the validation criteria.
16

17 ASSAY

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19 Carry out the assay of heparin (2.7.5). The estimated potency is not less than 90 per
20 cent and not more than 111 per cent of the stated potency. The confidence limits of the
21 estimated potency ($P = 0.95$) are not less than 80 per cent and not more than 125 per
22 cent of the stated potency.
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24 STORAGE

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26 In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof
27 container.
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29 LABELLING

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31 The label states:

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33 — the number of International Units per milligram;
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35 — the animal species of origin; |
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37 — where applicable, that the substance is suitable for use in the manufacture of parenteral
38 preparations.
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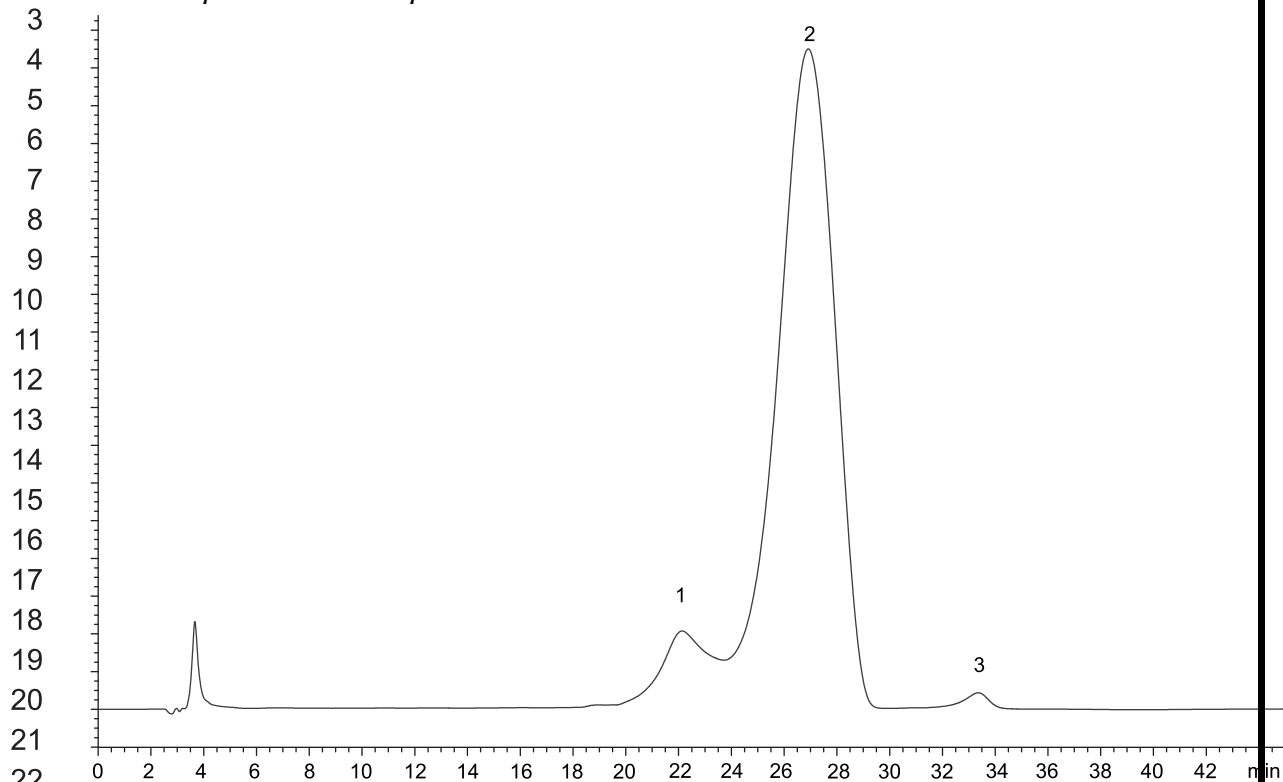
40 Reagents

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42 **Deuterated sodium trimethylsilylpropionate**. $C_6H_9^2H_4NaO_2Si$. (M_r 172.3). XXXXXXXX.
43 [24493-21-8]. Sodium 3-(trimethylsilyl)(2,2,3,3- 2H_4)propionate. TSP- d_4 .
44

45 *Degree of deuteration*: minimum 98 per cent.
46

47 White or almost white powder.

1 The following chromatogram is shown for information but will not be published in
2 the European Pharmacopoeia.



23 1. dermatan sulfate + chondroitin 2. heparin 3. over-sulfated chondroitin
24 sulfate sulfate sulfate

25 Figure 0332.-1.- Chromatogram for identification test C of heparin calcium: reference
26 solution (c) (chromatogram obtained after subtraction of the blank)

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The following chromatogram is shown for information but will not be published in the European Pharmacopoeia.

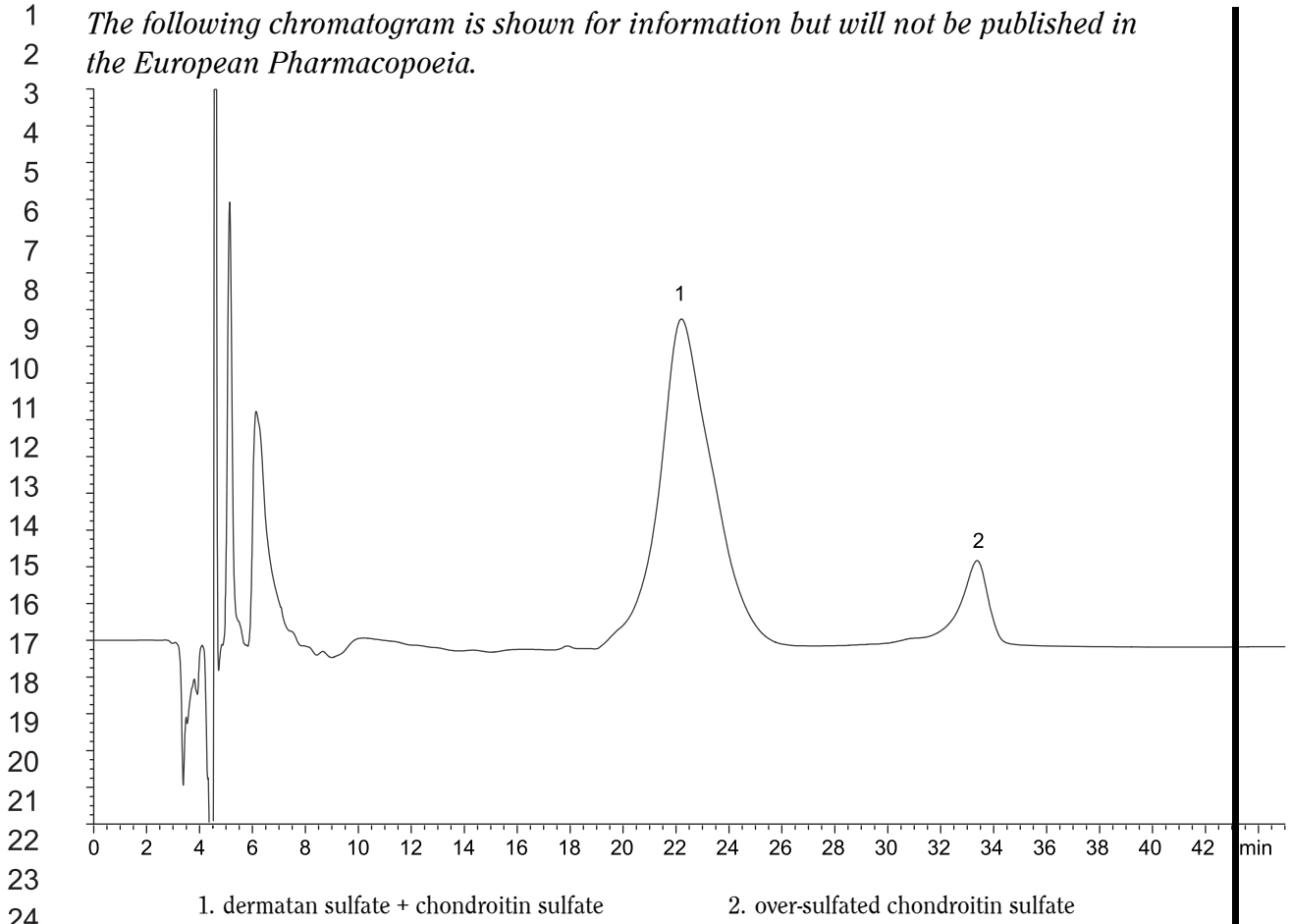


Figure 0332.-2.- Chromatogram for the test for related substances of heparin calcium: reference solution (e) (chromatogram obtained after subtraction of the blank)

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